Effects of mifepristone on invasive and metastatic potential of human gastric adenocarcinoma cell line MKN-45 in vitro and in vivo

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INTRODUCTION

Mifepristone is a progesterone receptor (PR) antagonist that has been widely used as the first-line drug for the termination of early pregnancy[1]. Recently, considerable studies[2,3] have proved that mifepristone exerts markedly anti-tumor effects on PR-positive tumor cells, such as breast cancer and ovarian cancer, without obvious side-effects and drug resistance. Its similar effect on PR-positive human gastric adenocarcinoma cell line MKN-45 was also demonstrated in our laboratory.

More interestingly, accumulating evidences show that embryo implantation and tumor metastasis share striking similarities in biological behaviors, such as cell adhesion[4], immune escape[5], angiogenesis[6] and tumor metastasis-related gene expression[7]. By this rational, there is an increasing interest in addressing the role of mifepristone, an agent against embryo implantation, in anti-tumor invasion and metastasis. Therefore, the present study was undertaken to further investigate the effects of mifepristone on the invasive and metastatic potential of MKN-45 cells in vitro and in vivo and its possible mechanisms.

MATERIALS AND METHODS

Cell culture and treatment

Human gastric adenocarcinoma cell line MKN-45 was obtained from Wuhan University Type Culture Collection (Wuhan, China), and maintained in phenol red-free RPMI1640 (Gibco BRL, Grand Island, NY) supplemented with 100 µL/L fetal bovine serum (Hyclone, Logan, UT), 10³ U/L penicillin and 100 µg/L streptomycin at 37 °C in a humidified atmosphere containing 5% CO₂ in air. When cells were grown to approximately 50% confluence, the medium was then replaced with serum-free RPMI1640. After 24 h, fresh media containing 5, 10, 20 µmol/L mifepristone (Sigma Chemical Co., St Louis, MO) were added, respectively. Control cells were treated with the same volumes of vehicle (ethanol). Unless otherwise stated, the cells were harvested after 96 h of incubation.

Adhesion assay

Each well in 96-well tissue culture plates was coated with 2 µg of Matrigel (Collaborative Research Inc., Bedford, MA) and allowed to dry in a laminar flow cabinet overnight at room temperature. After washed three times with PBS to remove excess and unbound Matrigel, the wells were blocked with 20 µL of a 20 mg/L bovine serum albumin (BSA, Sigma) solution in RPMI1640 medium for 1 h at 37 °C. Aliquots of 8×10⁴ cells, in 100 µL of serum-free RPMI1640 medium containing various concentrations of mifepristone, were added to each well and the cells were allowed to adhere for 1 h at 37 °C. When the incubation was completed, the wells were washed three times with PBS to remove unbound cells. Then, the remaining cells were continuously incubated with MTT solutions (40 µg/well) for 4 h at 37 °C, followed by treatment with 200 µL of dimethyl sulfoxide (DMSO) for 10 min. Finally, A570nm of each well was measured using an ELISA plate reader(Bio-Rad, USA). Results were expressed as the adhesive rate(%) that was calculated according to the
following formula: \((A_{570nm} \text{ of the adhered cells}/A_{570nm} \text{ of total cells}) \times 100\%\).

**Detection of integrin β3 by flow cytometry**

The harvested cells were fixed with 40 mg/L paraformaldehyde for 10 min, followed by treatment with 2 mg/L Triton X-100 for 10 min. After treatment with normal rabbit serum for 10 min to block non-specific binding, the cells were incubated for 1 h at 4 °C with mouse anti-human monoclonal antibody against β3 (Santa Cruz, USA), followed by treatment with FITC-conjugated goat anti-mouse IgG (Vector, Burlingame, CA) for 30 min at 4 °C. The percentage of positive cells was determined using the FACS Calibur flow cytometry (Becton & Dickinson) with an excitation wavelength of 488 nm.

**Migration assay**

Migration of MKN-45 cells was assayed in Transwell cell culture chambers with 6.5-mm-diameter polycarbonate membrane filters containing 8-µm-pore size (Becton Dickenson Labware, Bedford, MA). Fibroblast-conditioned medium, obtained from confluent NIH 3T3 cell cultures in serum-free RPMI 1640, was used as the chemoattractant and added to the lower wells of the chambers. Aliquots of 2×10^4 cells in 300 µL fresh medium containing various concentrations of mifepristone were seeded into the upper wells of the cell inserts. After 24 h of incubation at 37 °C, the non-migrating cells were removed from the upper surface of the membrane with a cotton swab. The cells on the lower surface of the membrane were fixed with ice-cold methanol and then stained with hematoxylin and eosin. The number of migrated cells was counted under a light microscope. Five random microscopic fields (×400) were counted per well and the mean was determined.

**Measurement of VEGF by ELISA**

The media were collected after 48 h for VEGF ELISA determinations as described below. The cells were taken through three freeze-thaw cycles, centrifuged and supernatant was collected for determination of protein concentration as described previously. VEGF levels in the cell culture media were measured using a Quantikine kit from R & D Diagnostics (Minneapolis, MN) using the procedure provided by the supplier. Human recombinant VEGF included in the kit was used to construct a standard curve and obtain absolute values of VEGF protein content. The values were then normalized to the total protein concentration in each dish.

** Xenografts of MKN-45 cells in nude mice**

Two×10^7 MKN-45 cells were subcutaneously xenografted in the right flank of 8-wk-old male BALB/c- nu/nu mice (Shanghai Experimental Animal Center, Chinese Academy of Sciences, China). When tumors reached a mean volume of 100 mm³, mice were randomly divided into two groups (8 mice in per group) and treated as the following. Mice in experiment group were administrated subcutaneously with mifepristone at the dose of 50 mg/kg·d, whereas mice in control were subcutaneously injected with saline every day. After 8 wk of treatment, lungs were harvested, and the weights of lungs were determined. The number of metastatic foci in lungs fixed with Bouin’s solution for 24 h was counted under a stereomicroscope. Meanwhile, the tumors were resected, fixed with 40 g/L formaldehyde in PBS, embedded in paraffin and sliced into 4-µm-thick sections for immunohistochemical analysis.

**Immunohistochemical staining for VEGF and MVD**

The expression of VEGF and MVD in harvested tumors was determined immunohistochemically using an avidin-biotin-peroxidase complex (ABC) kit (Vector Laboratories, USA). Unless otherwise stated, all steps were performed at room temperature. Briefly, after removal of wax, tissue sections were treated with 30 mL/L hydrogen peroxidase for 30 min to block endogenous peroxidase activity, and microwaved in 10 mmol/L citrate buffer for 10 min to retrieve antigens. After blocked with normal goat serum, rabbit anti-human polyclonal antibody against VEGF and factor VIII-related antigen (Santa Cruz Biotechnology, USA) at a 100-fold dilution were separately applied to sections and incubated overnight at 4 °C. This was followed by treatment with biotin-labeled goat anti-rabbit IgG for 60 min. The ABC complex was added and allowed to stand for 30 min. Sites of immunoreaction were visualized with 3', 3'-diaminobenzidine (DAB), followed by counterstaining with Mayer’s haematoxylin, if necessary. Between each step, the sections were washed three times with 100 mmol/L Tris-HCl buffer containing 0.1 mg/L Triton X-100. For a positive control, breast cancer tissue was used and the primary antibody was replaced by normal rabbit serum as a negative control.

**Statistical analysis**

Data were expressed as mean±SD. Statistical analysis was performed using the Student’s t test and chi-square test. \(P<0.05\) was considered statistically significant.

**RESULTS**

**Adhesion assay and expression of integrin β3**

MTT assay revealed that mifepristone dose-dependently inhibited the cell adhesion to artificial basement membrane, Matrigel. The adhesive rate of MKN-45 cells was 78.2±5.0%, 65.4±4.7%, 49.8±4.2% in mifepristone-treated group, and 85.6±6.3% in control group (\(P<0.05\)). To further explore whether the change of cell adhesion molecule expression in MKN-45 cells contributed to the inhibition, flow cytometry was used to detect the expression of integrin β3 in the cells. Results showed that the expression of integrin β3 in MKN-45 cells treated with 5, 10, 20 µmol/L mifepristone was significantly lower than that in control group (25.4±3.6%, 19.6±2.9%, 15.8±2.2% vs 31.8±4.1%, \(P<0.05\)).

**Migration assay**

The effect of mifepristone on the migration of MKN-45 cells was evaluated in the Transwell cell culture chambers. As shown in Figure 1, there was a significant decrease in the number of migrated MKN-45 cells contributed to the inhibition. Flow cytometry was used to compare with that in control cells (22.7±4.3 ng/g per liter, \(P<0.01\)).

**Expression of VEGF protein**

After treatment with 5, 10, 20 µmol/L mifepristone for 48 h, secreted VEGF protein in the cell culture media measured by ELISA assay, was 14.2±2.9, 8.9±3.1 and 5.4±2.1 ng/g per liter, respectively. There was a significant difference in VEGF expression as compared with that in the control cells (22.7±4.3 ng/g per liter, \(P<0.01\)).

**Xenografts of MKN-45 cells in nude mice**

During necropsy, lung metastasis of gastric cancer was found in 6 mice of the control group, and in 4 mice of mifepristone-treated group (Figure 2). As far as the number of metastatic foci in lungs was concerned, there was a significant difference in mifepristone-treated group as compared with control group (8±2 vs 18±7, \(P<0.05\)). In addition, the weights of xenografted tumors in nude mice treated with mifepristone (201±36 mg) were significantly lower than those in control group (298±54 mg, \(P<0.05\)). To evaluate the effects of mifepristone on MVD and VEGF expression in xenografted tumors, immunohistochemical
staining was performed using ABC method. Results revealed that VEGF was highly expressed in xenografted gastric cancer in nude mice (Figure 3A). After treatment with mifepristone for 8 wk, the expression of VEGF was markedly downregulated in the tumors (Figure 3B). In addition, MVD of xenografted tumors was (11.2±2.5)/400×visual field in mifepristone-treated group, and (29.8±7.6)/400×in the control group (P<0.01).

**Figure 1** Effect of mifepristone on migration of MKN-45 cells incubated in Transwell cell culture chambers for 24 h in the absence (A) or presence of 10 µmol/ L (B) and 20 µmol/ L (C) mifepristone (H.E, ×200).

**Figure 2** Metastatic foci in lungs of nude mice (arrow). After 24 h of fixation with Bouin’ s solution, lung tissues exhibited yellow, whereas the tumors exhibited white.

**Figure 3** Immunohistochemical detection of VEGF expression in xenografted gastric cancer tissues of nude mice treated for 8 wk without (A) or with (B) 50 mg/ kg·d mifepristone (ABC, ×100).

**DISCUSSION**

Although considerable evidence[9,10], both clinical and experimental, has demonstrated that mifepristone exerts markedly anti-proliferative effects on PR-positive malignant tumor cells, the role of mifepristone in anti-tumor invasion and metastasis, especially on gastric cancer, is poorly understood. In the present study, we demonstrated that mifepristone effectively inhibited the invasive and metastatic potential of human gastric adenocarcinoma cell line MKN-45 in vitro and in vivo through multiple mechanisms.

For tumor cells, increase of heterotypic adhesion to basement membrane and decrease of homotypic adhesion to the same cells have been defined as the critical event of tumor invasion that signals the initiation of metastatic cascade[11,12]. In the present study, heterotypic adhesion of MKN-45 cells to artificial basement membrane, Matrigel, was examined with MTT dye assay to stain the adhered cells. Results showed that mifepristone dose-dependently decreased the adhesive rate of MKN-45 cells. Moreover, the findings were further proved by the down- regulation of integrin β3 expression in the cells treated with mifepristone.

Integrin β3, one member of integrins superfamily, plays a fundamental role in tumor cell heterotypic adhesion to extracellular matrix and basement membrane, which has been found to be mediated through a specific arginine-glycine-aspartic acid (RGD) amino acid sequence[13]. Brakebusch et al.[14] reported that high expression of integrin β3 directly correlated with tumorigenicity and tumor progression. To further explore whether the change of integrin β3 expression in MKN-45 cells contributed to the inhibitory effect of mifepristone on cell adhesion, flow cytometry was performed. We found that mifepristone down-regulated the expression of integrin β3 in MKN-45 cells in a dose-dependent manner. The results were also demonstrated by the work of Li et al.[15], who reported that mifepristone significantly down-regulated the expression of integrin β3 in decidua and chorionic villi of early pregnancy, and the effect might be related with the anti-pregnancy mechanism of mifepristone. Taken together, the inhibitory
effect of mifepristone on heterotypic adhesion of MKN-45 cells may be associated with the down-regulation of integrin β3. Moreover, integrin β3 has been found to function not only as a cell adhesion molecule, but also as a signaling molecule for regulation of angiogenesis[16]. Therefore, down-regulation of integrin β3 expression in MKN-45 cells may be partially responsible for the inhibition of angiogenesis in the cells by mifepristone.

Continuous growth, invasion and metastasis of malignant tumors including human gastric cancer are dependent on angiogenesis factors regulated by peptide growth factors, of which VEGF is one of the most selective and potent[17]. Hyder et al.[19] reported that progesterin could induce the expression of VEGF mRNA and protein in human breast cancer cell line T47D, and this effect was blocked by the antiprogestin agent mifepristone. The finding suggests that mifepristone may be useful to inhibit proliferation and metastasis in some tumors by blocking VEGF production. Thus, there is a great interest in exploring the effect of mifepristone on the expression of VEGF in MKN-45 cells. After exposure of MKN-45 cells to various concentrations of mifepristone for 48 h, a dose-dependent decrease in the media levels of VEGF was observed. Furthermore, the results were further supported by animal experiments. Immunohistochemical analysis showed that mifepristone significantly down-regulated the expression of VEGF and MVD in xenografted gastric cancer in nude mice. Summarily, it seems reasonable to conclude that the inhibitory effect of mifepristone on the invasive and metastatic potential of MKN-45 cells is mediated partially via blocking VEGF production.

On the other hand, tumor cell migration was necessary at the initiation of metastatic cascade, when the tumor cells left the primary site and gained access to the circulation and also at the end of invasion, when they were entering the secondary site[19]. Theoretically, the decrease of tumor cell migration would contribute to the inhibition of tumor invasion and metastasis. In the study, the effect of mifepristone on the migration of MKN-45 cells was assessed in Transwell cell culture chambers. Results showed that a significant decrease in the number of migrated MKN-45 cells was estimated in Transwell cell culture. These findings, and metastatic potential of human MKN-45 gastric adenocarcinoma cells through inhibition of the heterotypic adhesion to basement membrane, cell migration and angiogenesis. These findings, linked to its anti-proliferative effects, indicate that mifepristone may be a beneficial agent for additional and complementary use in the management of gastric cancer.

REFERENCES


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