

Thymoma and Thymic Carcinoma

**Protocol applies to thymic epithelial tumors located
in any area of the mediastinum.**

*Protocol revision date: January 2004
No AJCC/UICC staging system*

Procedures

- Biopsy
- Resection

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The College of American Pathologists offers these protocols to assist pathologists in providing clinically useful and relevant information when reporting results of surgical specimen examinations of surgical specimens. The College regards the reporting elements in the “Surgical Pathology Cancer Case Summary (Checklist)” portion of the protocols as essential elements of the pathology report. However, the manner in which these elements are reported is at the discretion of each specific pathologist, taking into account clinician preferences, institutional policies, and individual practice.

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Summary of Changes to Checklist(s)

Protocol revision date: January 2004

No changes have been made to the data elements of the checklist(s) since the January 2003 protocol.

Surgical Pathology Cancer Case Summary (Checklist)

*Protocol revision date: January 2004
Applies to all epithelial thymic neoplasms
No AJCC/UICC staging system*

THYMOMA AND THYMIC CARCINOMA: Biopsy

Patient name:

Surgical pathology number:

Note: Check 1 response unless otherwise indicated.

MACROSCOPIC**Specimen Type**

- Fine-needle aspiration biopsy
 Computed tomography-guided needle biopsy
 Transthoracic needle biopsy
 Limited thoracotomy
 Other (specify): _____
 Not specified

Tumor Site

- Thymus
 Anterior mediastinum
 Middle mediastinum
 Posterior mediastinum
 Other (specify): _____
 Not specified

MICROSCOPIC**Histologic Type**

- Type A thymoma (epithelial, spindle cell, medullary)
- Type B thymoma, B1 (lymphocyte-rich, lymphocytic, predominantly cortical, organoid)
- Type B thymoma, B2 (cortical)
- Type B thymoma, B3 (epithelial, atypical, squamoid, well-differentiated thymic carcinoma)
- Type AB thymoma (mixed)
- Type C thymoma (thymic carcinoma), epidermoid keratinizing (squamous cell) carcinoma
- Type C thymoma (thymic carcinoma), epidermoid nonkeratinizing carcinoma/lymphoepithelioma-like carcinoma
- Type C thymoma (thymic carcinoma), sarcomatoid carcinoma
- Type C thymoma (thymic carcinoma), carcinosarcoma
- Type C thymoma (thymic carcinoma), clear cell carcinoma
- Type C thymoma (thymic carcinoma), basaloid carcinoma
- Type C thymoma (thymic carcinoma), mucoepidermoid carcinoma
- Type C thymoma (thymic carcinoma), papillary carcinoma
- Type C thymoma (thymic carcinoma), undifferentiated carcinoma
- Other (specify): _____
- Carcinoma, type cannot be determined

***Comment(s)**

* Data elements **with asterisks** are **not required** for accreditation purposes for the Commission on Cancer. These elements may be clinically important, but are not yet validated or regularly used in patient management. Alternatively, the necessary data may not be available to the pathologist at the time of pathologic assessment of this specimen.

Surgical Pathology Cancer Case Summary (Checklist)

*Protocol revision date: January 2004
Applies to all epithelial thymic neoplasms
No AJCC/UICC staging system*

**THYMOMA AND THYMIC CARCINOMA: Thymectomy,
Other Procedure**

Patient name:

Surgical pathology number:

Note: Check 1 response unless otherwise indicated.

MACROSCOPIC**Specimen Type**

- Cervical thymectomy
 Thoracotomy
 Video-assisted thoracotomy
 Other (specify): _____
 Not specified

***Specimen Size**

- *Greatest dimension: ____ cm
 *Additional dimensions: ____ x ____ cm

Tumor Site

- Thymus
 Anterior mediastinum
 Middle mediastinum
 Posterior mediastinum
 Other (specify): _____
 Not specified

Tumor Size

- Greatest dimension: ____ cm
 *Additional dimensions: ____ x ____ cm
 Cannot be determined (see Comment)

MICROSCOPIC**Histologic Type**

- Type A thymoma (epithelial, spindle cell, medullary)
 Type B thymoma, B1 (lymphocyte-rich, lymphocytic, predominantly cortical, organoid)
 Type B thymoma, B2 (cortical)
 Type B thymoma, B3 (epithelial, atypical, squamoid, well-differentiated thymic carcinoma)
 Type AB thymoma (mixed)
 Type C thymoma (thymic carcinoma), epidermoid keratinizing (squamous cell) carcinoma
 Type C thymoma (thymic carcinoma), epidermoid non-keratinizing carcinoma/lymphoepithelioma-like carcinoma
 Type C thymoma (thymic carcinoma), sarcomatoid carcinoma
 Type C thymoma (thymic carcinoma), carcinosarcoma
 Type C thymoma (thymic carcinoma), clear cell carcinoma
 Type C thymoma (thymic carcinoma), basaloid carcinoma
 Type C thymoma (thymic carcinoma), mucoepidermoid carcinoma
 Type C thymoma (thymic carcinoma), papillary carcinoma
 Type C thymoma (thymic carcinoma), undifferentiated carcinoma
 Other (specify): _____
 Carcinoma, type cannot be determined

Pathologic Staging

- Stage I: Grossly and microscopically encapsulated
 Stage IIa: Microscopic transcapsular invasion
 Stage IIb: Macroscopic capsular invasion
 Stage III: Macroscopic invasion of neighboring organs
 Stage IVa: Pleural or pericardial dissemination
 Stage IVb: Hematogenous or lymphatic dissemination
 Cannot be determined

Regional Lymph Nodes

- Cannot be assessed
 No regional lymph node metastasis
 Regional lymph node metastasis
 Specify: Number examined: ____
 Number involved: ____

Distant Metastasis

- Cannot be assessed
 Distant metastasis
 *Specify site(s), if known: _____

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Margins

- Cannot be assessed
- Margins uninvolved by tumor
Distance of tumor from closest margin: ____ mm
- Margin(s) involved by tumor
Specify margin(s): _____

Invasion of Pulmonary Parenchyma

- Cannot be assessed
- Absent
- Present
- Indeterminate

Pleural Invasion

- Cannot be assessed
- Absent
- Present
- Indeterminate

***Vascular (Small/Large Vessel) Invasion**

- * Absent
- * Present
- * Indeterminate

***Additional Pathologic Findings**

*Specify: _____

***Comment(s)**

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Background Documentation

Protocol revision date: January 2004

I. Biopsy

A. Clinical Information

1. Patient identification
 - a. Name
 - b. Identification number
 - c. Age (birth date)
 - d. Sex
2. Responsible physician(s)
3. Date of procedure
4. Date of specimen receipt in pathology laboratory
5. Previous/concurrent cytology or biopsy specimen (Note **A**)
6. Other relevant clinical information
 - a. History (eg, lung cancer, myasthenia gravis, previous diagnosis, treatment)
 - b. Imaging and laboratory findings (eg, computed tomography [CT] scan, magnetic resonance imaging [MRI], positron emission tomography [PET] scan, operative findings)
 - c. Clinical findings and diagnosis(es)
 - d. Previous or concurrent therapy, including dates (eg, surgery, radiation, chemotherapy, other)
 - e. Procedure(s) (eg, CT-guided needle biopsy, mediastinoscopic biopsy, limited thoracotomy)
 - f. Findings at procedures (eg, mediastinoscopy, limited thoracotomy)
 - g. Anatomic site(s) of specimen(s) (eg, thymus, anterior mediastinum, posterior mediastinum, middle mediastinum)
 - h. Other

B. Macroscopic Examination

1. Specimen
 - a. Unfixed/fixed (specify fixative)
 - b. Size (3 dimensions)
 - c. Descriptive features
 - d. Results of intraoperative consultation
2. Tissue submitted for microscopic evaluation
 - a. Entire specimen or selected samples
 - b. Frozen section tissue fragment(s) (unless saved for special studies)
3. Special studies (specify)

C. Microscopic Evaluation

1. Tumor, if present
 - a. Histologic type (Note **B**)
 - b. Extent of invasion, as appropriate
 - c. Vascular and lymphatic invasion
 - d. Perineural invasion
 - e. Other (specify)
2. Additional pathologic findings, if present
3. Status/results of special studies (specify)
4. Comments
 - a. Correlation with intraprocedural consultation, as appropriate
 - b. Correlation with other specimens, as appropriate
 - c. Correlation with clinical information, as appropriate

II. Resection

A. Clinical Information

1. Patient identification
 - a. Name
 - b. Identification number
 - c. Age (birth date)
 - d. Sex
2. Responsible physician(s)
3. Date of procedure
4. Date of specimen receipt in pathology laboratory
5. Previous/concurrent cytology or biopsy specimen (Note **A**)
6. Other relevant clinical information
 - a. History (eg, lung cancer, myasthenia gravis, previous diagnosis, treatment)
 - b. Imaging and laboratory findings (eg, CT scan, PET scan, operative)
 - c. Clinical findings and diagnosis(es)
 - d. Previous or concurrent therapy, including dates (eg, surgery, radiation, chemotherapy, other)
 - e. Procedure(s) (eg, thymectomy, cervical or mediastinal; thoracotomy; other)
 - f. Operative findings
 - g. Anatomic sites of specimen(s)
 - h. Other

B. Macroscopic Examination

1. Specimen
 - a. Organs/tissues received (documentation of extent of resection)
 - b. Unfixed/fixed (specify fixative)
 - c. Size of entire specimen (3 dimensions)
 - d. Weight
 - e. External aspect (Note **C**)
 - (1) encapsulated, invasive borders
 - (2) attached tissue (eg, parietal pleura, pericardium, diaphragm, chest wall with or without ribs, other)
 - f. Documentation of specific areas marked by surgeon
 - g. Results of intraoperative consultation
2. Tumor
 - a. Location
 - (1) thymus
 - (2) other (eg, paraesophageal, peribronchial, pericardial, others)
 - b. Size (3 dimensions)
 - c. Descriptive features
 - (1) color
 - (2) shape
 - (3) circumscription
 - (4) cavitation
 - (5) other (eg, necrosis, hemorrhage)
 - d. Extent of invasion
 - (1) structures involved by invading tumor, including vessels and nerves
 - e. Additional tumors, if present
 - (1) size (range)
 - (2) number
 - (3) location
 - f. Margins (specify distance from closest approach of tumor)
 - g. Additional pathologic findings, if present

- h. Regional lymph nodes in specimen
 - (1) location
 - (2) number
 - (3) description
 - i. matted
 - ii. gross metastasis
 - iii. size of largest lymph node containing tumor
 - iv. extranodal extension of tumor
- i. Sections of tissue for microscopic evaluation (Note **D**)
 - (1) tumor (at least 1 section per centimeter per maximum tumor diameter)
 - (2) tumor interface with adjacent tissues
 - (3) tumor invading adjacent tissues; adjacent tissues containing tumor
 - (4) tumor capsule (capsule should be histologically sampled in areas of capsular disruption; otherwise, multiple random capsular sections should be made)
 - (5) margins
 - (6) frozen section tissue fragment(s) (unless saved for special studies)
 - (7) specific areas designated by surgeon
 - (8) areas with additional pathologic findings
 - (9) other organs(s), tissues
- 3. Special studies (specify)
- 4. Photography

C. Microscopic Evaluation

- 1. Tumor
 - a. Histologic type (Note **B**)
 - b. Site/location
 - c. Transcapsular invasion, extent
 - d. Vascular invasion (arteriolar or venous)
 - e. Lymphatic invasion
 - f. Perineural invasion
 - g. Adjacent structures/organs
- 2. Margins
 - a. Presence
 - b. Margin width (in millimeters)
- 3. Status of area(s) marked by surgeon
- 4. Additional pathologic findings
- 5. Non-neoplastic tissues from site of origin (eg, thymus)
- 6. Regional lymph nodes included in main specimen
 - a. Total number examined
 - b. Number involved by tumor
 - c. Size of the largest metastasis
 - d. Extracapsular extension present or absent
 - e. Metastases to other organs
- 7. Additional pathologic findings, if present
- 8. Results of special studies (specify) (Note **E**)
- 9. Stage (Note **F**)
- 10. Comments
 - a. Correlation with intraoperative consultation, as appropriate
 - b. Correlation with other specimens, including cytology, as appropriate
 - c. Correlation with clinical information, as appropriate

Explanatory Notes

A. Cytologic Findings

Pathologists should indicate the nature and clinical significance of any cytologic abnormality as specifically as possible. Fine-needle aspiration biopsies of mediastinal masses have a reasonably high yield for the diagnosis of thymoma and carcinomas. Cell blocks are particularly helpful, as they can be used for immunocytochemical studies.

B. Histologic Type

Levine and Rosai have classified tumors of the thymic epithelium as encapsulated and invasive (malignant) thymomas and thymic carcinoma.¹⁻⁴ In general, the Cancer Committee of the College of American Pathologists provides guidelines solely for malignant neoplasms, such as invasive thymomas and thymic carcinomas. Although encapsulated thymomas are benign neoplasms in the vast majority of patients, because they can recur locally in a small number of patients and because distant metastases have been reported in rare patients, they are included in this protocol. Levine and Rosai have subclassified both encapsulated and invasive thymomas, based on histopathologic features, into epithelial, lymphocytic, and mixed lymphocytic and epithelial.¹ More recently, Marino and Muller-Hermelink have proposed a histological classification of thymomas designating them as cortical, mixed (common, with cortical predominance, with medullary predominance), and medullary.⁵ This classification is widely used in Europe but is not accepted by most American pathologists because of diagnostic reproducibility problems.⁶ Recently, the World Health Organization (WHO) proposed the following grouping of thymomas and thymic carcinomas^{7,8}:

Type A Thymoma (spindle cell, medullary)

Type B Thymoma

B1 thymoma (lymphocyte-rich, lymphocytic, predominantly cortical, organoid)

B2 thymoma (cortical)

B3 thymoma (epithelial, atypical, squamoid, well-differentiated thymic carcinoma)

Type AB Thymoma (mixed)

Type C Thymoma (thymic carcinoma)

Epidermoid keratinizing (squamous cell) carcinoma

Epidermoid nonkeratinizing carcinoma/lymphoepithelioma-like carcinoma

Sarcomatoid carcinoma

Carcinosarcoma

Clear cell carcinoma

Basaloid carcinoma

Mucoepidermoid carcinoma

Papillary carcinoma

Undifferentiated carcinoma

Type A thymomas are composed of epithelial cells with oval or spindle-shaped nuclei and few lymphocytes. This tumor type corresponds to the designation of epithelial thymomas of the Levine and Rosai classification scheme. Types B1 and B2 thymomas are composed of large numbers of lymphocytes admixed with a fewer epithelial cells. These tumors correspond to the designation of lymphocytic thymomas of the Levine and Rosai scheme. Type B3 thymomas correspond to thymomas with atypical histology, which were not clearly defined by Levine and Rosai. Type AB thymomas correspond to mixed lymphoepithelial thymomas of the Levine and Rosai scheme. Thymic carcinomas include a variety of malignant cytologic features and are designated as Type C thymomas.⁹⁻¹⁷

C. Designation of Areas Suspicious for Invasion

Areas of adherence of the mediastinal mass to other mediastinal structures may be the only indication of tumor capsular penetration and hence the only indication of tumor malignancy. Surgeons should be strongly encouraged to refrain from incising the tumor capsule prior to examination by a pathologist; incisions result in tissue retraction and can compromise margin assessment. Uncertainties regarding the nature and degree of capsular adherence should be discussed with the surgeon(s) who removed the tumor. Any areas of macroscopic adherence or otherwise deemed suggestive of invasion should be marked by the surgeon postexcision and histologically sampled.

D. Number of Sections to Submit

The number of sections submitted varies with the size and character of the specimen and the nature of the underlying neoplastic process. Tumors with a heterogeneous cut surface should be sampled more thoroughly. The capsule of thymomas should be sectioned more thoroughly than the central area of the tumors. One section per centimeter of tumor largest diameter is recommended for most neoplasms.

E. Special Studies in Mediastinal Lesions

Thymomas and thymic carcinomas usually require immunocytochemistry or, less frequently, electron microscopy to establish a diagnosis. The types of special studies that must be obtained vary with the histologic appearance of the tumor as it appears on initial examination. Immunostains for keratin are helpful in distinguishing between thymomas and lymphoid lesions. In selected cases, the use of immunohistochemistry for CD1a and terminal deoxynucleotidyl transferase (TdT) may be helpful in defining the cortical thymocyte phenotype of thymoma, as distinguished from the typical peripheral T-cell phenotype of tumor-infiltrating lymphocytes associated with other tumors. Immunostains for human chorionic gonadotropin, placental alkaline phosphatase, carcinoembryonic antigen, and α -fetoprotein are helpful in differentiating among thymic carcinomas and mediastinal germ cell tumors.

F. Staging of Thymic Epithelial Neoplasms

No TNM protocol has been proposed by the American Joint Committee on Cancer (AJCC) or the International Union Against Cancer (UICC) for the staging of thymic epithelial neoplasms. The scheme developed by Masaoka as modified by Koga et al is frequently used for staging¹⁸⁻²¹:

Stage I	Grossly and microscopically completely encapsulated (including microscopic invasion into the capsule)
Stage IIa	Microscopic transcapsular invasion
Stage IIb	Macroscopic capsular invasion into thymic or surrounding fat, or grossly adherent but not breaking through mediastinal pleura or pericardium
Stage III	Macroscopic invasion of neighboring organs (eg, pericardium, lung, great vessels, others)
Stage IVa	Pleural or pericardial dissemination
Stage IVb	Hematogenous or lymphatic dissemination

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